Escherichia coli O157 in Bovine Feces and Surface Water Streams in a Beef Cattle Farm of Argentina

José D. Tanaro,¹ Gerardo A. Leotta,^{2,3} Liliana H. Lound,¹ Lucía Galli,^{2,3} Mercedes C. Piaggio,¹ Carolina C. Carbonari,² Santiago Araujo,¹ and Marta Rivas²

Abstract

Shiga toxin-producing *Escherichia coli* (STEC) is an important foodborne pathogen, and ruminants are recognized as the main natural reservoir. The purposes of this study were to detect *E. coli* O157 in bovine feces and surface water in a beef cattle farm of Gualeguaychú, Argentina; to characterize the isolates; and to establish the clonal relatedness by pulsed-field gel electrophoresis. Between September 2005 and November 2006, 288 samples of bovine feces and 79 samples of water troughs were studied. *E. coli* O157 was detected by immunomagnetic separation and polymerase chain reaction as screening techniques. The rfb_{O157} gene was detected in 3.8% of the 288 fecal samples and in 17.7% of the 79 water samples. The stx gene was detected in all rfb_{O157} -positive fecal samples and in 5.1% of water samples. Eleven *E. coli* O157 strains isolated from bovine fecal samples and eight from water samples were characterized. The most frequent stx genotype identified was stx_1 and $stx_{2c(vh-a)}$. Twelve (63.2%) strains harbored $fliC_{H7}$, *eae*, and *ehx*A genes. Using pulsed-field gel electrophoresis with the enzyme XbaI, a total of eight patterns with at least 72.1% similarity were identified among the 19 strains. The patterns of 15 strains. No genetic correlation was established between the bovine and water STEC strains detected. The prevalence of STEC O157:H7 established in the herd studied was higher than that previously reported for Argentine grazed cattle.

Introduction

S HIGA TOXIN-PRODUCING *Escherichia coli* (STEC) are zoonotic agents that are widely distributed around the world and are important foodborne pathogens. The prototype strain is *E. coli* O157:H7. Cattle are considered the main animal reservoir for STEC O157:H7, and the first isolates in cattle were described in Argentine calves in 1977 (Caprioli *et al.*, 2005). STEC infection can cause bloody diarrhea and hemolytic uremic syndrome (HUS). Argentina has the highest HUS rate globally—13.9/100,000 children under age 5—and *E. coli* O157:H7 has been related to almost 75% of the cases (Rivas *et al.*, 2006). The purposes of this study were to detect *E. coli* O157, especially serotype O157:H7, in bovine feces and water in a beef cattle farm of Gualeguaychú, Argentina; to characterize the isolates; and to establish the clonal relatedness by pulsed-field gel electrophoresis (PFGE).

Materials and Methods

Between September 2005 and November 2006, 288 swabs of rectal feces from healthy animals (Rice *et al.*, 2003) and 79 water

samples (Spira and Ahmed, 1981) were collected. Sample size was calculated using the Epi Info software (version 6.0), taking into account an estimated frequency of 10% and a precision of 1.5% at 95% confidence level. The estimated prevalence was based on a 0.5% prevalence previously reported (Meichtri et al., 2004), which was multiplied by a factor of 20 because of an increase in sensitivity resulting from the use of the immunocapture system. Rectal swabs were placed in 100 mL of modified EC broth (Biokar Diagnostics, Beauvois, France) with 20 mg/L of sodium novobiocin (mEC+N; Sigma Chemical, St. Louis, MO). The water samples were taken with modified Moore swabs from two streams, one inside the study area (200 acres), accessible for the animals, and the other downhill out of the field. The swabs were placed in flasks with 100 mL of $_{m}EC+N.$ Enrichment broths were incubated for 24 h at 42°C and processed by immunomagnetic separation (Dynal, Compiàgne, France); the immunoconcentrates were streaked on two media: O157:H7 ID™ (bioMérieux, Marcy l'Etoile, France), and MacConkey sorbitol agar (Biokar) supplemented with 2.5 mg/L of potassium tellurite and 0.05 mg/L of cefixime (bioMérieux). After incubation, confluent growth zone and

¹School of Food Science, National University of Entre Ríos, Gualeguaychú, Argentina.

²Branch of Physiopathogenesis, Department of Bacteriology, National Institute of Infectious Diseases—ANLIS "Dr. Carlos G. Malbrán," Buenos Aires, Argentina.

³National Council of Scientific and Technical Research (CONICET), Buenos Aires, Argentina.

individual colonies were screened for stx_1 , stx_2 , and rfb_{O157} genes by multiplex polymerase chain reaction (PCR) (Leotta *et al.*, 2005).

The isolates were characterized by standard biochemical tests (Ewing, 1986) and genotypic techniques (Leotta *et al.*, 2008). The antimicrobial susceptibility was carried out by the Kirby–Bauer method following the recommendations of the Clinical and Laboratory Standards Institute. *Xba*I-PFGE was performed according to the Centers for Disease Control and Prevention protocol (CDC, 2007). *Bln*I (Promega, Madison, WI) was used as second enzyme.

Results and Discussion

Eleven (3.8%) out of the 288 fecal samples were rfb_{O157} positive. Two samples were detected in autumn and nine in winter. STEC strains were isolated from all rfb_{O157}-positive samples. Four (36%) of them harbored stx_2 genes, and seven (64%) carried stx_1 and stx_2 sequences, simultaneously (Fig. 1). All strains were β -D-glucuronidase negative, cytotoxic on Vero cells, showed enterohemolysis production, and were susceptible to ampicillin, ampicillin-sulbactam, piperacillin, cephalothin, cefoxitin, cefuroxime, cefotaxime, ceftazidime, nalidixic acid, ciprofloxacin, trimethoprim-sulfamethoxazole, nitrofurantoin, gentamicin, chloramphenicol, fosfomycin, and tetracycline. The PCR-restriction fragment length polymorphism genotyping method showed that seven strains harbored stx_1 and $stx_{2c(vh-a)}$ genes and four carried only $stx_{2c(vh-a)}$ sequence. All strains were positive for *fliC_{H7}, eae*, and *ehxA* genes. Fourteen (17.7%) out of the 79 water samples were rfb_{O157} positive. Three samples were detected in spring, four in summer, one in autumn, and six in winter. However, E. coli O157 strains were isolated from eight (10.1%) water samples. Seven strains had none of the genes for virulence factors. The STEC strain isolated from a water sample showed the same phenotypic characteristics as the bovine isolates and harbored stx_2 and $stx_{2c(vh-a)}$, fliC_{H7}, eae, and ehxA genes. XbaI-PFGE was able to subtype the 19 E. coli O157 strains, generating eight different patterns with 16-21 discernible fragments, ranging approximately from 36 to 582 kb in molecular weight. Fifteen strains with indistinguishable profiles were grouped into four clusters, and four strains showed unrelated XbaI-PFGE patterns (Fig. 1). Cluster I (AREXHX01.0625) included seven toxigenic bovine strains isolated in May and July 2006. Six of them presented the same BlnI-PFGE pattern, and the remaining strain differed in only one band, with 97.4% similarity. Cluster II (AREXHX01.0626) grouped three toxigenic bovine strains, isolated at the same sampling date (July 17, 2006), as some of the strains grouped in cluster I. They were differentiated into two BlnI-PFGE profiles, with only one band difference and 97.3% similarity. Cluster III (AREXHX01.0296) included three nontoxigenic strains isolated from water. Two of them were isolated in August 2006, in two different streams. One strain of this cluster differed in the BlnI-PFGE profile, with only one band difference and 96% similarity. It was ampicillin resistant. Cluster IV (AREXHX01.0435) included two nontoxigenic water strains isolated in the same stream in October and November 2006. These strains were indistinguishable by *Bln*I-PFGE.

A previous report from Argentina confirmed that calves harbored STEC O157:H7, with a prevalence of 0.5% (Meichtri et al., 2004). In this study, the prevalence of E. coli O157:H7 in fecal samples of grazed cattle was 3.8%. These isolates were recovered during autumn and winter in 2006. Both precipitation (240.5 mm) and temperature (10.6-19.8°C) in May, June, and July 2006 were higher than normal (Argentine National Weather Service). In water samples, E. coli O157 could only be isolated from 57% of PCR-positive samples but only one isolate was toxigenic. This strain harbored stx_2 and $stx_{2c(yh-a)}$ genes, which is the prevalent genotype in bloody diarrhea and HUS cases (Rivas et al., 2006). It presented the AREXHX01.0298 profile not previously included in the E. coli O157 Argentine Database. However, Shiga toxin-negative E. coli O157 strains detected in water could be able to acquire stx genes as described by Wetzel and LeJeune (2007). The eight strains recovered from water showed five different XbaI

PFGE-Xbal	x) (TOI 1	.5%-1.5%) (H>00% S>0.0%) (K5%-5	18.3%]							
ا ج	Ę	PFGE-Xbal	PFGE-BIN	Xbal-PFGE pattern	Bhil-PFGE pattern	Origir	n Isolation dat	te Place	Serotype	Genetic profile
				AREXHX01.0625	AREXHA26.0098	в	05-02-2006	Raising area	O157:H7	eae / ehxA / stx1 / stx2c (vh-a)
				AREXHX01.0625	AREXHA26.0098	Ð	07-10-2006	Raising area	O157:H7	eae / ehxA / stx1 / stx2c (vh-a)
				AREXHX01.0625	AREXHA26.0098	в	07-10-2006	Raising area	O157:H7	eae / ehxA / stx1 / stx2c (vh-a)
	- Li			AREXHX01.0625	AREXHA26.0098	в	07-10-2006	Raising area	O157:H7	eae / ehxA / stx1 / stx2c (vh-a)
				AREXHX01.0625	AREXHA26.0098	в	07-17-2006	Raising area	O157:H7	eae/ehxA/stx1/stx2c(vh-a)
	-			AREXHX01.0625	AREXHA26.0099	в	07-17-2006	Raising area	O157:H7	eae / ehxA / stx1 / stx2c (vh-a)
				AREXHX01.0625	AREXHA26.0098	в	07-17-2006	Raising area	O157:H7	eae / ehxA / stx1 / stx2c (vh-a)
				AREXHX01.0314		в	04-18-2006	Raising area	O157:H7	eae / ehxA / stx2c (vh-a)
Ы				AREXHX01.0626	AREXHA26.0103	в	07-17-2006	Raising area	O157:H7	<i>eae / ehx</i> A / <i>stx2c</i> (vh-a)
				AREXHX01.0626	AREXHA26.0104	в	07-17-2006	Raising area	O157:H7	eae / eħxA / stx2c (vh-a)
-	- Ľ			AREXHX01.0626	AREXHA26.0103	в	07-17-2006	Raising area	O157:H7	eae / eħxA / stx2c (vh-a)
Π	Ĩ			AREXHX01.0296	AREXHA26.0101	w	08-13-2006	Stream I	0157:HNT	nb 0157
				AREXHX01.0296	AREXHA26.0102	W	08-13-2006	Stream II	0157:HNT	nb 0157
				AREXHX01.0296	AREXHA26.0101	W	07-04-2006	Stream I	0157:HNT	nb 0157
L				AREXHX01.0295		w	08-13-2006	Stream I	0157:HNT	nb 0157
۲ـــــــــــــــــــــــــــــــــــــ			i	AREXHX01.0298		w	11-08-2006	Stream II	O157:H7	eae / ehxA / stx2 / stx2c (vh-a)
	i.			AREXHX01.0435	AREXHA26.0100	w	11-08-2006	Stream II	0157:HNT	nb 0157
<u> </u>		VARCES HELENER		AREXHX01.0435	AREXHA26.0100	w	10-05-2006	Stream II	0157:HNT	nb 0157
				AREXHX01.0635		w	07-04-2006	Stream I	0157:HNT	nb 0157

FIG. 1. Phenotypic and genotypic characterization and clonal relatedness, into 4 clusters, of 19 *Escherichia coli* O157 strains isolated from cattle and water in a beef cattle farm, Gualeguaychú, Argentina, 2005–2006.

E. COLI 0157 IN BOVINE AND WATER IN ARGENTINA

profiles, whereas the 11 bovine strains showed only three *XbaI* profiles. This would indicate that there is more diversity in the aquatic environment.

In conclusion, in the herd studied we found (a) a higher prevalence of STEC O157:H7 than that previously reported for Argentine grazed cattle, (b) the toxigenic strain recovered from water was genetically different from the toxigenic animal strains, and (c) during the study period, strains from both sources, animal and environment, showed different virulence profiles. However, other studies should be conducted to better understand the risk of STEC transmission in Argentina, considering bovine and environmental reservoirs or both.

Acknowledgments

The authors thank Sergio Sosa-Estani for his helpful suggestions, and Lucía Isturiz for critical reading of the manuscript.

Disclosure Statement

No competing financial interests exist.

References

- Caprioli A, Morabito S, Brugère H, *et al.* Enterohaemorrhagic *Escherichia coli:* emerging issues on virulence and modes of transmission. Vet Res 2005;36:289–311.
- [CDC] Centers for Disease Control and Prevention. One-Day (24–28 h) Standardized Laboratory Protocol for Molecular Subtyping of *Escherichia coli* O157:H7 by Pulsed Field Gel Electrophoresis. Atlanta, GA: Centers for Disease Control and Prevention, 2007.
- Ewing WH. Identification of Enterobacteriaceae, 4th edition. New York: Elsevier, 1986.
- Leotta GA, Chinen I, Epszteyn S, et al. Validación de una técnica de PCR múltiple para la detección de Escherichia coli productor

de toxina Shiga. Rev Argent Microbiol 2005;37:1-11. (In Spanish.)

- Leotta GA, Miliwebsky E, Chinen I, Espinosa EM, Azzopardi K, Tennant SM, Robins-Browne RM, and Rivas M. Characterization of Shiga toxin-producing *Escherichia coli* O157 strains isolated from humans in Argentina, Australia and New Zealand. BMC Microbiol 2008;8:46–53.
- Meichtri L, Miliwebsky E, Gioffré A, *et al.* Shiga toxin-producing *Escherichia coli* in healthy young beef steer from Argentina: prevalence and virulence properties. Int J Food Microbiol 2004;96:189–198.
- Rice DH, Sheng HQ, Wynia SA, *et al.* Rectoanal mucosal swab culture is more sensitive than fecal culture and distinguishes *Escherichia coli* O157:H7-colonized cattle and those transiently shedding the same organism. J Clin Microbiol 2003;41:4924–4929.
- Rivas M, Miliwebsky E, Chinen I, *et al.* Epidemiología del Síndrome Urémico Hemolítico en Argentina. Diagnóstico del agente etiológico, reservorios y vías de transmisión. Medicina Buenos Aires 2006;66:27–32. (In Spanish.)
- Spira WM and Ahmed QS. Gauze filtration and enrichment procedures for recovery of *Vibrio cholerae* from contaminated waters. Appl Environ Microbiol 1981;42:730–733.
- Wetzel AN and LeJeune JT. Isolation of *Escherichia coli* O157:H7 strains that do not produce Shiga toxin from bovine, avian and environmental sources. Lett Appl Microbiol 2007;45: 504–507.

Address correspondence to: José D. Tanaro, M.D. Facultad de Bromatología Universidad Nacional de Entre Ríos Presidente Perón 64 Gualeguaychú 2820 Argentina

E-mail: jdtanaro@fb.uner.edu.ar

This article has been cited by:

- 1. Pianciola Luis, Chinen Isabel, Mazzeo Melina, Miliwebsky Elizabeth, González Gladys, Müller Constanza, Carbonari Carolina, Navello Mariano, Zitta Eugenia, Rivas Marta. 2014. Genotypic characterization of Escherichia coli O157:H7 strains that cause diarrhea and hemolytic uremic syndrome in Neuquén, Argentina. *International Journal of Medical Microbiology*. [CrossRef]
- José D. Tanaro, Lucía Galli, Liliana H. Lound, Gerardo A. Leotta, Mercedes C. Piaggio, Carolina C. Carbonari, Kinue Irino, Marta Rivas. 2012. Non-O157:H7 Shiga Toxin–Producing Escherichia coli in Bovine Rectums and Surface Water Streams on a Beef Cattle Farm in Argentina. *Foodborne Pathogens and Disease* 9:10, 878-884. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF with Links]
- 3. Robert W. Derlet, John R. Richards, Charles R. Goldman. 2012. Does Above-Normal Precipitation Reduce the Impact of Mountain Cattle Grazing on Watershed Algae and Bacteria?. *Water Quality, Exposure and Health* 4:2, 105-112. [CrossRef]
- 4. Beatriz A. D'Astek, Lourdes L. del Castillo, Elizabeth Miliwebsky, Claudia Carbonari, Pablo M. Palladino, Natalia Deza, Isabel Chinen, Eduardo Manfredi, Gerardo A. Leotta, Marcelo O. Masana, Marta Rivas. 2012. Subtyping of Escherichia coli O157:H7 Strains Isolated from Human Infections and Healthy Cattle in Argentina. *Foodborne Pathogens and Disease* 9:5, 457-464. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF with Links]