

## ORIGINAL RESEARCH

## Effect of $\gamma$ radiation processing on fungal growth and quality characteristics of millet grains

Nagat S. Mahmoud<sup>1</sup>, Sahar H. Awad<sup>1</sup>, Rayan M. A. Madani<sup>1</sup>, Fahmi A. Osman<sup>1</sup>, Khalid Elmamoun<sup>2</sup> & Amro B. Hassan<sup>1</sup>

<sup>1</sup>Environment and Natural Resource and Desertification Research Institute (ENDRI), National Center for Research, PO Box 6096, Khartoum, Sudan

<sup>2</sup>Sudanese Atomic Energy Commission (SAEC), Khartoum, Sudan

### Keywords

Antinutritional factors, fungal growth, germination, millet, protein solubility, radiation

### Correspondence

Amro B. Hassan, Environment and Natural Resource and Desertification Research Institute (ENDRI), National Center for Research, PO Box 6096, Khartoum, Sudan.  
Tel: +249912244812;  
Fax: +249188463441;  
E-mail: amrobabiker@yahoo.com

### Funding Information

No funding information provided.

Received: 18 June 2015; Revised: 24 August 2015; Accepted: 16 September 2015

*Food Science & Nutrition* 2016; 4(3): 342–347

doi: 10.1002/fsn3.295

### Abstract

The aim of this study was to evaluate the effect of gamma radiation processing of millet grains on fungal incidence, germination, free fatty acids content, protein solubility, digestible protein, and antinutritional factors (tannin and phytic acid). The grains were exposed to gamma radiation at doses 0.25, 0.5, 0.75, 1.0, and 2.0 kGy. Obtained results revealed that radiation of millet grains at a dose level higher than 0.5 kGy caused significant ( $P < 0.05$ ) reduction on the percentage of fungal incidence and the free fatty acid of the seeds, while, no significant change in the germination capacity was observed of the grains after radiation. Additionally, the radiation process caused significant ( $P < 0.05$ ) reduction on both tannins and phytic acid content and gradual increment on in vitro protein digestibility of the grains. On the other hand, the treatments significantly ( $P < 0.05$ ) increased the protein solubility of the grains. Obtained results indicate that gamma irradiation might improve the quality characteristics of millet grains, and can be used as a postharvest method for disinfestations and decontamination of millet grains.

## Introduction

Pearl millet (*Pennisetum glauca* L.) is considered to be the staple food for most people in Asia and Africa. It is considered as a good source of needed elements (Abdalla et al. 1998). However, it contains high amounts of antinutrients such as tannin and phytic acids which reduce its nutritional value (Abdel Rahaman et al. 2007). During postharvest storage, millet is susceptible to attack by a variety of insects and microorganisms. This infestation cause physical losses and reduces the nutritional value of grains which leads to the loss of the economic value of stored grain. Moreover, infestation with the insects result in contamination with dead insect's bodies and their products, as well as fungal growth that favor the spread of *Aspergillus flavus*, a mold which produces aflatoxin (Rees 2004).

Generally, chemical fumigants are used to disinfest grains (Arthur 1996), however, continuous application of these pesticides have a negative impact either on the environment or human health (Cherry et al. 2005). Therefore, the industry has been forced to explore nonchemical alternatives. One possible alternative is the application of gamma irradiation. Radiation processing is considered to be a safe alternative to chemical methods, enhance quality and nutritional characteristics of stored products as well as maintain its shelf-life. It has shown great promise in accomplishing disinfestations and decontamination of food and agricultural products (Loaharanu 1994; Fombang et al. 2005). Besides disinfection criteria, gamma radiations enhance the nutritional value of grains and improves the functional properties of its flours (Rahma and Mostafa 1988; Dario and Salgado 1994; Dogbevi et al. 2000). Furthermore, it has been reported that gamma radiation

causes a significant reduction in antinutrients and enhances the nutritional quality of grains (Hassan *et al.* 2013; Osman *et al.* 2014). Although, application of gamma radiation has many advantages over other physical methods, however, the application of this technology in the industry is limited. Therefore, further research is needed to improve their efficiency to reach the reasonable usage stage and in controlling stored-grain pests as well as to improve the nutritive value of stored products.

Thus, in the present study, gamma radiation was applied as a preserving method to investigate its efficiency in controlling fungal growth and enhancement of quality characteristics of millet grains.

## Material and Methods

### Sample preparation

Millet grains were cleaned manually and freed from broken seeds and impurities, and then stored in plastic bags at 4°C during the study.

### Radiation treatments

Radiation processing was done at Kaila irradiation processing unit, Sudanese Atomic Energy Corporation (SAEC). About 250 g of millet grains packed in polyethylene bags were irradiated, using a  $\gamma$ -ray  $^{60}\text{Co}$  radiator. The seeds were evenly exposed to radiation doses 0.25, 0.5, 0.75, 1.0, and 2.0 kGy with a dose rate of 33 Gy/min. Unirradiated seeds (0 kGy) served as control.

### Fungal culture and incidence

The fungal incidence and the colony formation unit per gram (cfu/g) of treated and untreated samples was determined after platted on double strength Sabaroud Dextrose Agar and incubated at 25°C for 5 days according to standard methods (AOAC 1995).

### Determination of germination

The germination of grains was determined according to the international Seed Testing Association (ISTA 2006). Twenty five seeds were platted on filter paper in a Petri dish and saturated with distilled water. The plates were incubated at  $25 \pm 2^\circ\text{C}$  for 7 days.

### Determination of free fatty acid content

Free fatty acid of millet was determined according to Aibara *et al.* (1986) cited by Zhao *et al.* (2007) with slight modification. About 25 mL of ethanol was added

to 5 g of millet flour. After shaking, the mixture was filtrated and additional 25 mL ethanol was added. The filtrate was titrated with 0.1 N KOH, using phenolphthalein (3%) as indicator. Flour acidity was calculated as mg KOH required neutralizing free fatty acid from one gram grain on dry matter basis.

### Determination of tannins and Phytic acid content

Tannins content of grains was estimated according to Price *et al.* (1978). Phytic acid content was determined by the method described by Wheeler and Ferrel (1971).

### Crude and digestible protein determination

The crude protein was determined following the Kjeldahl method described by AOAC (1995). The digestible protein was determined by the procedure of Maliwal (1983) cited from Monjula and John (1991).

$$\text{Protein digestibility \%} = \frac{\text{digestible protein}}{\text{total protein}} \times 100$$

### Protein solubility

Soluble protein solubility was determined in millet grains after extracted by water, using the method described by Hagenmaier (1972).

### Statistical analysis

All data were the average of triplicates. Data were analyzed using one-way analysis of variance (ANOVA). Significant differences were calculated ( $P < 0.05$ ), using least significant difference (LSD).

## Results and Discussion

### Effect of radiation process on the fungi incidence and germination rate

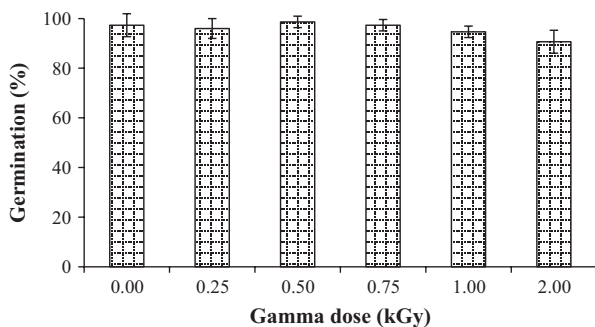
Table 1 presents the percentage of fungal incidence and colony formation (cfu/g) in raw and treated millet grains. Before radiation, the fungal incidence was found to be 100% in raw grains. Radiation of seeds up to 0.50 kGy caused no significant reduction in fungal incidence, however, radiation of grains at higher doses 0.75, 1.0 and 2.0 kGy sharply decreased the fungal incidence to 21.3, 18.7, and 5.3%, respectively. Similarly, the effectiveness of gamma radiation in reducing the formation of fungi was observed. Prior to radiation, the colony formation was found to be  $5.3 \times 10^4$  cfu/g, where as it was decreased to  $2.1 \times 10^4$ ,  $2.1 \times 10^4$ ,  $4 \times 10^2$ ,  $3 \times 10^2$  and  $3 \times 10$  cfu/g

**Table 1.** Effect of gamma irradiation on fungal growth (%) and colony formation (cfu/g) in pearl millet. Error bars indicate the standard deviation ( $n = 3$ ). Values not sharing a common superscript are significantly ( $P < 0.05$ ) different.

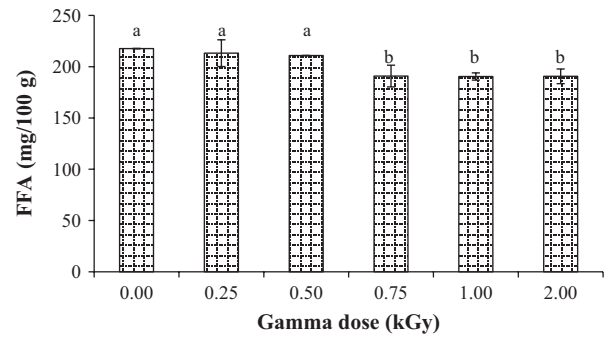
Gamma dose (kGy)	Fungal incidence (%)	Colony formation (cfu/g)
0.0	100 ± (0.000) <sup>a</sup>	5.3 × 10 <sup>4</sup>
0.25	100 ± (0.000) <sup>a</sup>	2.1 × 10 <sup>4</sup>
0.50	100 ± (0.000) <sup>a</sup>	2.1 × 10 <sup>4</sup>
0.75	21.3 ± (9.238) <sup>b</sup>	4.0 × 10 <sup>2</sup>
1.0	18.7 ± (2.309) <sup>b</sup>	3.0 × 10 <sup>2</sup>
2.0	5.3 ± (2.309) <sup>c</sup>	3.0 × 10 <sup>1</sup>

after radiation treatment at doses of 0.25, 0.50, 0.75, 1.0, and 2.0 kGy, respectively (Table 1). Similar observation on walnut kernel was reported by Al-Bachir (2004), who found that application of gamma radiation at doses of 0.5, 1.5, and 2 kGy reduced the fungal load on walnut kernels. Furthermore, it was reported that doses of 1.5 and 3.5 kGy reduced the number of fungi in many raw fruits and vegetables (Aziz and Moussa 2004). Hilmy et al. (1995) concluded that radiation process peanuts with doses up to 1 kGy inhibit the incidence of mycelium and toxins secretion. Reduction in the fungal incidence rate in millet grains after radiation might be due to high sensitivity of the fungus and mold to gamma radiation, since the radiation process causes direct and indirect damage to the DNA (Refai et al. 1996; McNamara et al. 2003).

Germination test is comparatively assessing the quality losses of grain after treatments. It is directly associated with various characteristics of grain quality (Beckett and Morton 2003). As shown in Figure 1, no significant change in seed germination rate after treatments was observed. Maximum decrease in germination rate of millet (90.7%) was observed with 2 kGy treatment. Obtained results were in accordance with El-Naggar and Mikhael (2011), who found that the germination capacity of wheat grains was not changed after radiation at dose up to 1 kGy. Moreover, the results of Melki and Marouani (2009) showed that



**Figure 1.** Effect of gamma irradiation on germination rate in pearl millet. Error bars indicate the standard deviation ( $n = 3$ ).

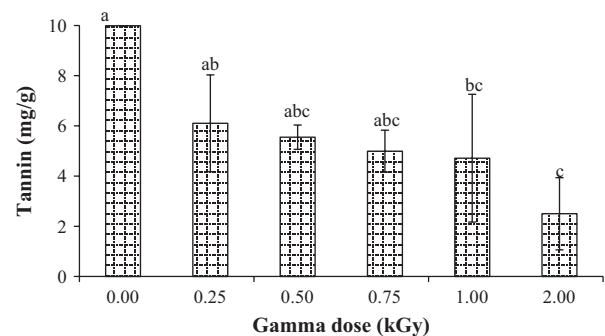


**Figure 2.** Effect of gamma irradiation on free fatty acids (FFA) in pearl millet. Error bars indicate the standard deviation ( $n = 3$ ). Values not sharing a common superscript are significantly ( $P < 0.05$ ) different.

there was no significant change in germination capacity in wheat after radiation.

### Effect of radiation process on free fatty acids (FFA) content

Figure 2 presents the free fatty acids (FFA) content in mg/g in millet grains for the control and radiated samples. The FFA content of millet flour was found to be 217.7 mg/100 g prior to the radiation treatment. After radiation at dose levels of 0.75, 1.0, and 2.0 kGy, it is clearly observed that the FFA content of millet grains significantly ( $P < 0.05$ ) reduced to 190.9, 190.6, and 190.5 mg/100 g, respectively. Significant reduction on FFA content might be due to lipase activity reduction in treated grains, which result in dropping the FFA formation. Pankaj et al. (2013) demonstrated that the radiation treatment significantly reduced the lipase activity in wheat germ. Therefore, the results indicated that gamma radiation is an effective method for stabilization and to extend the shelf life of grains, since free fatty acids content is an index of the rancidity and contributes to the development of off-flavor and off- odors in oil during storage.

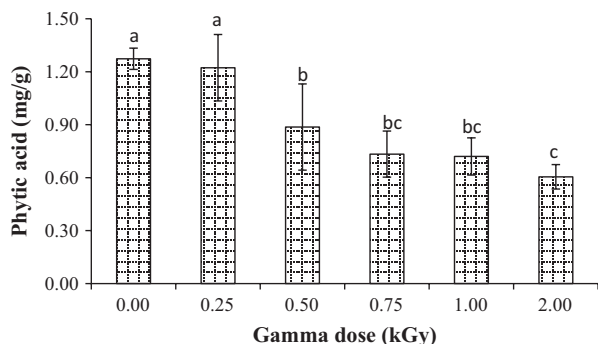


**Figure 3.** Effect of gamma irradiation on tannin content in pearl millet. Error bars indicate the standard deviation ( $n = 3$ ). Values not sharing a common superscript are significantly ( $P < 0.05$ ) different.

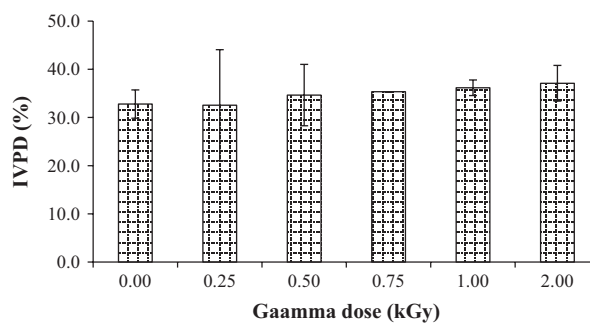
### Effect of radiation process on tannin and phytic acid content

As illustrated in Figure 3, the tannin content of millet grains was found to be 9.99 mg/g prior to radiation process. Tannin content of the examined grains presented a dose-dependent decrease. It was clearly observed that radiation significantly ( $P < 0.05$ ) reduced tannin content of millet grains. The reduction in tannin content was found to be 38.9, 44.4, 50, and 52.8, and 74.9% when millet grains were irradiated at dose 0.25, 0.50, 0.75, 1.0, and 2.0 kGy, respectively, compared to control one. These findings are in agreement with those stated by several researchers. Hassan et al. (2009) concluded that radiation process significantly reduced tannins content of sorghum and maize grains. Similar observation was reported by Pinn (1992) who stated that radiation of white beans at dose levels 2, 4, 6, 8, 10, 15, and 20 kGy followed by cooking significantly reduced tannins content. Moreover, El-Niely (2007) found that the tannin content of legume seeds decreased after radiation treatments. Decrease in tannin content might be result of chemical degradation by the action of free radicals formed by the radiation.

On the other hand, before radiation treatment, the phytic acid of millet grains was found to be 1.27 mg/g (Fig. 4). After radiation, phytic acid of millet grains significantly ( $P < 0.05$ ) decreased. The level of reduction increased with an increase in radiation dose. Decreases in phytic acid were 3.9, 29.9, 42.5, 43.3, and 52.8%, at dose 0.25, 0.50, 0.75, 1.0, and 2.0 kGy, respectively. This reduction might be the result of the action of free radicals, since they are able to cleave to the phytate ring (De Boland et al. 1975). Obtained results were in agreement with El-Niely (2007) who stated that radiation after processing significantly ( $P \leq 0.05$ ) decreased the level of phytic acid of legumes and cereal grains.



**Figure 4.** Effect of gamma irradiation on phytic acid content in pearl millet. Error bars indicate the standard deviation ( $n = 3$ ). Values not sharing a common superscript are significantly ( $P < 0.05$ ) different.

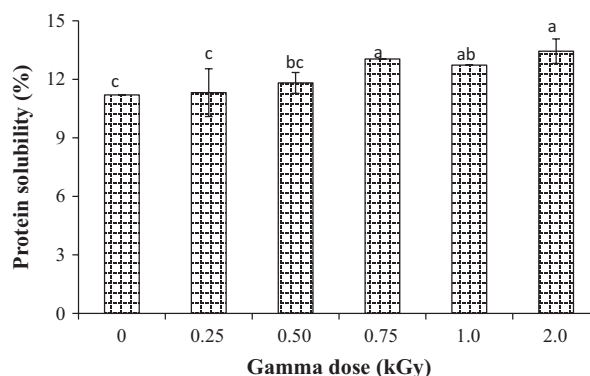


**Figure 5.** Effect of gamma irradiation on in vitro protein digestibility (%) in pearl millet. Error bars indicate the standard deviation ( $n = 3$ ).

### Effect of radiation process on in vitro protein digestibility (IVPD) and protein solubility

Figure 5 summarizes the in vitro protein digestibility (IVPD) of millet before and after radiation. The IVPD of untreated seeds was found to be 32.8%. Radiation process of the seeds caused a minor increment of the IVPD and was increased as the dose was increased. Increment in protein digestibility might be due to the reduction in the antinutrients particularly tannin content of grains as reported by Hassan et al. (2009). Since disulphide and hydrogen bonds are involved in stabilizing protein structure, their breaking can result in loss of conformational or structural integrity that exposed additional peptide bonds, thus enhancing proteolysis. Irradiation can cause change in their protein structure that enhances denaturation of the protein and hence improve its digestibility (Koppelman et al. 2005).

Protein solubility is doubtless the most important function among the functional properties of proteins. Data in Figure 6 showed that the protein solubility of raw



**Figure 6.** Effect of gamma irradiation on protein solubility in pearl millet. Error bars indicate the standard deviation ( $n = 3$ ). Values not sharing a common superscript are significantly ( $P < 0.05$ ) different.

millet was found to be 11.20%. The protein solubility of millet was increased significantly ( $P < 0.05$ ) to 11.32, 11.82, 13.04, 12.74, and 13.44% after radiation at 0.25, 0.50, 0.75, 1.0, and 2.0 kGy, respectively. Increase in protein solubility after radiation is likely due to the high proteolytic activity during radiation, which may lead to hydrolysis of the stored proteins.

## Conclusion

The obtained results revealed that gamma irradiation processing of millet grains up to 2 kGy significantly reduced the fungal incidence and free fatty acids content of the grains. On the other hand, it caused a decrease in the amount of antinutrients namely, tannin and phytic acid and gradually increase the *in vitro* protein digestibility and protein solubility of the grains. According to these results, therefore, gamma radiation can be applied as a safe postharvest method in order to extend the shelf life of millet grains.

## Conflict of Interest

None declared.

## References

- Abdalla, A. A., A. H. El Tinay, B. E. Mohamed, and A. H. Abdalla. 1998. Proximate composition, starch, phytate and mineral contents of 10 pearl millet genotypes. *Food Chem.* 63:243–246.
- Abdel Rahaman, S. M., H. B. ElMaki, W. H. Idris, A. B. Hassan, E. E. Babiker, and A. H. El Tinay. 2007. Antinutritional factors content and hydrochloric acid extractability of minerals in pearl millet cultivars as affected by germination. *Int. J. Food Sci. Nutr.* 58:6–17.
- Aibara, S., I. A. Ismail, H. Yamashita, and H. Ohta. 1986. Changes in rice bran lipids and fatty acids during storage. *Agric. Biol. Chem.* 50:665–673.
- Al-Bachir, M. 2004. Effect of gamma irradiation on fungal load, chemical and sensory characteristics of walnut (*Juglans regia*). *J. Stored Prod. Res.* 40:355–362.
- AOAC. 1995. Official methods of analysis of Association of Official Analytical Chemists, 16th ed. Association of Official Analytical Chemists, Washington, DC.
- Arthur, F. H. 1996. Grain protectants: current status and prospects for the future. *J. Stored Prod. Res.* 32:293–302.
- Aziz, N. H., and L. A. A. Moussa. 2004. Reduction of fungi and mycotoxins formation in seeds by gamma-radiation. *J. Food Safety* 24:109–127.
- Beckett, S. J., and R. Morton. 2003. Mortality of *Rhyzopertha dominica* (F.) at grain temperatures ranging from 50°C to 60°C obtained at different rates of heating in a spouted bed. *J. Stored Prod. Res.* 39:313–332.
- Cherry, A. J., P. Abalo, and K. Hell. 2005. A laboratory assessment of the potential of different strains of the entomopathogenic fungi *Beauveria bassiana* (Balsamo) Vuillemin and *Metarhizium anisopliae* (Metschnikoff) to control *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae) in stored cowpea. *J. Stored Prod. Res.* 41:295–309.
- Dario, A. C., and J. M. Salgado. 1994. Effect of thermal treatments on the chemical and biological value of irradiated and non-irradiated cowpea bean (*Vigna unguiculata* L.Walp.) flours. *Plant Food Hum. Nutr.* 46:181–186.
- De Boland, A. R., G. B. Garner, and B. L. O'Dell. 1975. Identification and properties of 'phytate' in cereal grains and oil-seed products. *J. Agric. Food Chem.* 23:1186–1189.
- Dogbevi, M. K., C. Vachon, and M. Lacroix. 2000. Effect of gamma irradiation on the microbiological quality and on the functional properties of proteins in dry red kidney beans (*Phaseolus vulgaris*). *Radiat. Phys. Chem.* 57:265–268.
- El-Naggar, S. M., and A. A. Mikhael. 2011. Disinfestation of stored wheat grain and flour using gamma rays and microwave heating. *J. Stored Prod. Res.* 47:191–196.
- El-Niely, H. F. G. 2007. Effect of radiation processing on antinutrients, *in vitro* protein digestibility and protein efficiency ratio bioassay of legume seeds. *Radiat. Phys. Chem.* 76:1050–1057.
- Fombang, E. N., J. R. N. Taylor, C. M. F Mbofung, and A. Minnaar. 2005. Use of  $\gamma$  irradiation to alleviate the poor protein digestibility of sorghum porridge. *Food Chem.* 91:695–703.
- Hagenmaier, R. 1972. Water binding of some purified oil seed proteins. *J. Food Sci.* 37:965–966.
- Hassan, A. B., G. A. M. Osman, M. A. Rushdi, M. M. Eltayeb, and E. E. Diab. 2009. Effect of gamma irradiation on the nutritional quality of maize cultivars (*Zea mays*) and sorghum (*Sorghum bicolor*) grains. *Pak. J. Nutr.* 8:167–171.
- Hassan, A. B., E. E. Diab, N. S. Mahmoud, R. A. A. Elagib, M. A. H. Rushdi, and G. A. M. Osman. 2013. Effect of radiation processing on *in vitro* protein digestibility and availability of calcium, phosphorus and iron of groundnut. *Radiat. Phys. Chem.* 91:200–203.
- Hilmy, N., R. Chosdu, and A. Matsuyama. 1995. The Effect of Humidity after gamma-irradiation on aflatoxin B production of *A. flavus* in ground nutmeg and peanut. *Radiat. Phys. Chem.* 46:705–711.
- International seed testing Association, ISTA. 2006. International rules for seed testing. Seed science and technology. ISTA, Basserdorf, Switzerland.
- Koppelman, S., W. F. Nieuwenh, M. Gaspari, L. M. J. Knippels, A. H. Pennincs, E. F. Knol, et al. 2005. Reversible denaturation of Brazil nut 2S albumins

- and implications and destabilization on digestion by pepsin. *J. Agric. Food Chem.* 53:123–131.
- Loaharanu, P. 1994. Food irradiation in developing countries: a practical alternative. *IAEA Bulletin.* 36:30–35.
- Maliwal, B. P. 1983. In vitro method to assess the nutritive value of leaf concentrate. *J. Agric. Food Chem.* 31:315–319.
- McNamara, N. P., H. I. J. Black, N. A. Beresford, and N. R. Parekh. 2003. Effects of acute gamma irradiation on chemical, physical and biological properties of soils. *Appl. Soil Ecol.* 24:117–132.
- Melki, M., and A. Marouani. 2009. Effects of gamma rays irradiation on seed germination and incidence of hard wheat. *Environ. Chem. Lett.* 8:307–331.
- Monjula, S., and E. John. 1991. Biochemical changes and in vitro protein digestibility of endosperm of germinating *Dolichos lablab*. *J. Sci. Food Agric.* 55:229–233.
- Osman, A. M., A. B. Hassan, G. A. M. Osman, N. Salih, M. A. H. Rushdi, E. E. Diab, et al. 2014. Effects of gamma irradiation and/or cooking on nutritional quality of faba bean (*Vicia faba* L.) cultivars seeds. *J. Food Sci. Technol.* 51:1554–1560.
- Pankaj, K. J., V. B. Kudachikar, and K. Sourav. 2013. Lipase inactivation in wheat germ by gamma irradiation. *Radiat. Phys. Chem.* 86:136–139.
- Pinn, A. B. R. O. 1992. Efeitos das radiações gama sobre adisponibilidade do ferro em feijões (*Phaseolus vulgaris*) (129 p). São Paulo: Dissertação (Mestrado). Faculdade de Ciências Farmacêuticas Universidade de São Paulo.
- Price, M. L., S. Van Socoyoc, and L. G. Butter. 1978. A critical evaluation of the vanillin reaction as an assay for tannin in sorghum grain. *J. Agric. Food Chem.* 26:1214–1218.
- Rahma, E. H., and M. M. Mostafa. 1988. Functional properties of peanut flour as affected by different heat treatments. *J. Food Sci. Technol.* 25:11–15.
- Rees, D. 2004. *Insects of stored products*. CSIRO publishing, Collingwood, VIC, Australia.
- Refai, M. K., N. H. Aziz, F. El-Far, and A. A. Hassan. 1996. Detection of ochratoxin produced by *A. ochraceus* in feedstuffs and its control by gamma radiation. *Appl. Radiat. Isot.* 47:617–621.
- Wheeler, E. I., and R. E. Ferrel. 1971. Methods for phytic acid determination in wheat and wheat fractions. *Cereal Chem.* 48:312–320.
- Zhao, S., S. Xiong, C. Qiu, and Y. Xu. 2007. Effect of microwaves on rice quality. *J. Stored Prod. Res.* 43:496–502.