



BRIEF REPORT

Serotypes, virulence profiles and *stx* subtypes of Shigatoxigenic *Escherichia coli* isolated from chicken derived products



Mónica Z. Alonso, Alejandra Krüger, Marcelo E. Sanz, Nora L. Padola*,
Paula M.A. Lucchesi

Laboratorio de Inmunoquímica y Biotecnología, Centro de Investigación Veterinaria de Tandil (CIVETAN), CONICET, CICPBA, Facultad de Ciencias Veterinarias, UNCPBA, Tandil, Buenos Aires, Argentina

Received 16 December 2015; accepted 12 April 2016

Available online 22 September 2016

KEYWORDS

STEC;
Chicken;
Serotypes;
Virulence factors;
stx subtypes

Abstract Shigatoxigenic *Escherichia coli* (STEC) is a foodborne pathogen that causes hemolytic uremic syndrome (HUS) and the consumption of chicken products has been related to some HUS cases. We performed a non-selective isolation and characterization of STEC strains from retail chicken products. STEC isolates were characterized according to the presence of *stx*₁, *stx*₂, *eae*, *saa* and *ehxA*; *stx* subtypes and serotypes. Most of them carried *stx*₂, showing subtypes associated with severe human disease. Although reported in other avian species, the *stx*_{2f} subtype was not detected. The isolates corresponded to different serotypes and some of them, such as O22:H8, O113:H21, O130:H11, O171:H2 and O178:H19, have also been identified among STEC isolated from patients suffering from diarrhea, hemorrhagic colitis, HUS, as well as from cattle. Considering the virulence profiles and serotypes identified, our results indicate that raw chicken products, especially hamburgers sold at butcheries, can be vehicles for high-risk STEC strains.

© 2016 Asociación Argentina de Microbiología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

PALABRAS CLAVE

STEC;
Pollo;
Serotipos;
Factores de
virulencia;
Subtipos de *stx*

Serotipos, perfiles de virulencia y subtipos de *stx* en *Escherichia coli* productor de toxina Shiga aislados de productos de pollo

Resumen *Escherichia coli* productor de toxina de Shiga (STEC) es un patógeno transmitido por alimentos que causa el síndrome urémico hemolítico (SUH). Algunos casos de SUH están relacionados con el consumo de productos de pollo. Se realizó el aislamiento no selectivo y la caracterización de cepas STEC provenientes de productos de pollo atendiendo a la presencia de *stx*₁, *stx*₂, *eae*, *saa* y *ehxA*, subtipos de *stx* y serotipos. La mayoría de los aislamientos

* Corresponding author.

E-mail address: nlpadola@vet.unicen.edu.ar (N.L. Padola).

<http://dx.doi.org/10.1016/j.ram.2016.04.009>

0325-7541/© 2016 Asociación Argentina de Microbiología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

portaba *stx*₂ y subtipos de *stx* asociados con enfermedades graves en humanos. Aunque se ha detectado en otras especies aviares, el subtipo *stx*_{2f} no se encontró. Se detectaron diferentes serotipos, entre ellos O22:H8, O113:H21, O130:H11, O171:H2 y O178:H19, también identificados como STEC aislados de pacientes con diarrea, colitis hemorrágica y SUH, y de ganado bovino. Teniendo en cuenta los perfiles de virulencia y los serotipos identificados, nuestros resultados indican que los productos de pollo crudos, especialmente las hamburguesas que se venden en las carnicerías, pueden ser vehículos de cepas STEC de alto riesgo.

© 2016 Asociación Argentina de Microbiología. Publicado por Elsevier España, S.L.U. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

Shigatoxigenic *Escherichia coli* (STEC) is a foodborne pathogen of public health importance that causes diarrhea, hemorrhagic colitis (HC) and hemolytic uremic syndrome (HUS) in humans. The main virulence factors of STEC are Shiga toxins (Stx1 and Stx2), which inhibit protein synthesis by inactivating ribosome function⁸. The Stx1 group is more homogenous than Stx2 since it includes only three subtypes. In contrast, a great number of subtypes have been identified for Stx2. The *stx* subtypes have been differently associated with HUS⁶. In addition to Shiga toxins, STEC can synthesize the adhesin intimin (encoded by *eae*), an enterohemolysin (EhxA), and an autoagglutinating protein (Saa) in some *eae*-negative strains, among other virulence factors⁸.

Different STEC serogroups have been identified in strains isolated from humans suffering from gastrointestinal disease. Five STEC serogroups (O26, O103, O111, O145, O157) are considered to be the "top five" serogroups most frequently associated with severe human disease in the European Union, and two others (O45 and O121) are also regarded as the most pathogenic ones in the USA. The serotype most frequently associated with outbreaks and sporadic cases of severe disease is O157:H7; however, more than 50% of all STEC infections are attributed to non-O157 strains³.

STEC transmission occurs through the consumption of contaminated food or water, direct contact with animals or their environments, and person-person contact⁸. With regard to food, the consumption of chicken products has been related to HUS cases, but most of the studies performed on this kind of products have been focused only on the detection of STEC O157:H7^{2,13}. Therefore, the aim of this study was to perform a non-selective isolation and characterization of STEC strains from retail chicken products.

Samples analyzed in the present study corresponded to 10 giblets and 54 chicken hamburgers previously identified as *stx*-positive by Alonso et al.¹ in a screening of 300 giblets and 300 chicken hamburgers. Peptone water cultures were stored at -70°C with 20% (v/v) glycerol. To isolate the STEC strains, an aliquot of each *stx*-positive culture, was streaked on MacConkey agar plates and incubated at 37°C for 24 h¹¹. Individual colonies were analyzed by a multiplex PCR to detect *stx*₁, *stx*₂, *eae*, *saa* and *ehxA* genes with the PCR protocol and primers described by Paton and Paton¹². Amplification products were electrophoresed in 2% agarose gels and stained with ethidium bromide. Only one colony was further characterized except when colonies with different virulence profiles were detected by this multiplex PCR.

As several samples were contaminated with *Proteus*, subsequent cultures were streaked repeatedly on cysteine lactose electrolyte deficient agar (CLED) to obtain pure colonies of *E. coli*. Afterwards, the absence of *Proteus* was verified by culture on a non-selective medium such as trypticase soy agar (TSA).

The O-antigens were determined by the microagglutination technique, and H antigens were determined by the tube agglutination technique using antisera provided by the Laboratorio de Referencia de *E. coli* (LREC) (Lugo, Spain) as described by Fernández et al.⁵.

To subtype *stx*₁ and *stx*₂ genes, PCR-restriction fragment length polymorphism (RFLP) assays were used⁹. In addition, a monoplex PCR described by Schmidt et al.¹⁴ was used to detect the *stx*_{2f} subtype.

Twenty-three STEC isolates were recovered from 54 *stx*-positive cultures of chicken hamburgers and only one isolate was obtained from 10 *stx*-positive giblet samples (Table 1). It was not possible to obtain any STEC isolate from some *stx*-positive samples although up to 200 colonies from those samples were analyzed.

Isolates carrying only the *stx*₂ gene predominated over the strains carrying both *stx*₁ and *stx*₂ or only *stx*₁, a similar trend to studies from other countries that detected *stx*₂ and not *stx*₁ in STEC isolated from chicken meat. This finding is important considering that Stx2 is more cytotoxic than Stx1⁸, and is associated with high virulence in humans⁶.

None of the STEC isolates carried the *eae* gene but some of them harbored the *saa* gene. STEC isolates that were *saa*-positive and *eae*-negative, belonging to serotypes O91:H21 and O113:H21 have been isolated from human patients with HUS⁸. Noticeably, in the present study O113:H21 isolates positive for *saa* were found in 3 samples and also harbored the *stx*_{2EDL933} subtype which has been associated with severe human disease.

Five virulence profiles could be determined by the multiplex PCR described by Paton and Paton¹², with *stx*₂ being the predominant profile (62.5%), followed by *stx*₂ *ehxA* *saa* and *stx*₁ *stx*₂ *ehxA* *saa* (17 and 12.5%, respectively). Furthermore, when the *stx* subtypes were also considered, 9 virulence profiles could be determined (Table 1).

With regard to STEC from chicken and derived products, there are few studies which identified the *stx* subtypes, and furthermore, these studies were focused exclusively on the characterization of O157:H7 strains². In the present study, all *stx*₁-positive isolates possessed the *stx*_{1EDL933} subtype, which has been associated with HUS cases and predominates in *stx*₁-positive isolates from cattle and meat products⁹. As

Table 1 Serotypes and virulence genes of STEC isolated from chicken products.

Number of isolates	Sample	Origin	Serotype	Virulence genes
1	Hamburger	Butchery	O22:H8	<i>stx</i> ₂ EDL933
1	Hamburger	Butchery	O91:H14	<i>stx</i> ₁ EDL933 <i>ehxA</i>
1	Hamburger	Butchery	O91:H40	<i>stx</i> ₂ EDL933 <i>ehxA</i>
3	Hamburger	Butchery	O113:H21 ^{a,b}	<i>stx</i> ₂ EDL933 <i>ehxA saa</i>
2	Hamburger	Butchery	O117:H7	<i>stx</i> ₂ vhb
2	Hamburger	Butchery	O130:H11 ^b	<i>stx</i> ₁ EDL933 <i>stx</i> ₂ vhb <i>ehxA saa</i>
1	Hamburger	Butchery	O130:H11	<i>stx</i> ₁ EDL933 <i>stx</i> ₂ EDL933 <i>ehxA saa</i>
1	Hamburger	Butchery	O153:H28	<i>stx</i> ₂ vhb
1	Hamburger	Butchery	O160:H40	<i>stx</i> ₂ EDL933
1	Hamburger	Butchery	O171:H2	<i>stx</i> ₂ O118
1	Hamburger	Butchery	O171:H2 ^a	<i>stx</i> ₂ vha
1	Hamburger	Butchery	O178:H19	<i>stx</i> ₂ vha
1	Hamburger	Butchery	ONT:H2	<i>stx</i> ₂ O118
1	Giblet	Butchery	ONT:H8	<i>stx</i> ₂ EDL933 <i>ehxA saa</i>
2	Hamburger	Butchery	ONT:H40	<i>stx</i> ₂ vhb
1	Hamburger	Poultry shop	ONT:H-	<i>stx</i> ₂ O118
3	Hamburger	Butchery	ONT:H-	<i>stx</i> ₂ vhb

^a One O113:H21 isolate and one O171:H2 isolate were obtained from the same sample.

^b One O113:H21 isolate and one O130:H11 isolate were obtained from the same sample.

far as we know, this is the first report about *stx*₁ subtypes in chicken samples. For the *stx*₂ gene, different subtypes were detected, but no more than one *stx*₂ subtype was identified within the same isolate. The *stx*₂vhb subtype was the most prevalent, accounting for 43.5% of the isolates, followed by *stx*₂EDL933 (8 isolates, 33%), *stx*₂O118 (3 isolates, 12%) and *stx*₂vha (2 isolates, 8%). This data is important because *stx*₂vhb, *stx*₂EDL933 and *stx*₂vha have been frequently associated with HUS cases⁶. The *stx*₂O118 subtype predominates in STEC strains from sheep, is rarely found in cattle, and, in contrast to our results, it is usually found associated with other *stx* subtypes in strains isolated from cattle and foods⁹.

Although *stx*_{2f}-positive strains have been isolated from avian species (pigeons), and Etoh et al.⁴ reported the isolation of a STEC strain harboring this subtype from a patient that had eaten raw chicken, we did not detect this subtype in any of the chicken samples.

STEC isolates belonging to O157:H7 were not detected in any of the samples, in agreement with the results obtained by other researchers who did not find this serotype in raw chicken meat and carcasses, even though they used selective methods for the isolation¹⁵. Indeed, there are only few studies that report the presence of STEC O157:H7 in chicken meat^{2,13}.

Several non-O157 serotypes were isolated from chicken products in the present study (Table 1). Some of the serotypes, such as O22:H8, O113:H21, O130:H11, O171:H2 and O178:H19, have also been isolated from patients suffering from diarrhea, HC or HUS, highlighting the importance of these findings⁷. Two hamburgers presented STEC isolates belonging to two different serotypes (O113:H21 and O171:H2 in one sample, and O113:H21 and O130:H11 in the other).

A comparison was made between STEC isolated in the present study and STEC isolates from cattle and derived products in Argentina. Noticeably, some serotypes such as O91:H14, O117:H7, O113:H21, O130:H11, O171:H2 and O178:H19 were present in both groups of strains,

and also *stx*₂ predominated over *stx*₁^{5,10,11}. Furthermore, some of the isolates belonging to serotypes such as O113:H21, O117:H7, O171:H2 and O178:H19 harbored the same virulence genotype and *stx* subtype as the isolates obtained from cattle, ground beef and evisceration tray samples in other studies^{9,10}.

In conclusion, the characterization of the STEC isolates in terms to serotype, virulence profile and *stx* subtype performed in the present study shows that chicken hamburgers can carry STEC strains that are potentially pathogenic to humans. Moreover, most of the isolates obtained from hamburgers presented the same serotype and genotype as the STEC strains recovered from cattle and derived meat products in our country.

Ethical responsibilities

Protection of human and animal subjects. The authors declare that no experiments were performed on humans or animals for this study.

Confidentiality of data. The authors declare that no patient data appear in this article.

Right to privacy and informed consent. The authors declare that no patient data appear in this article.

Conflict of interest

The authors declare that they have no conflicts of interest.

Acknowledgments

Authors thank María Rosa Ortiz for her technical assistance. This work was supported by grants from SECAT-UNICEN, CONICET and FONCYT. M. Z. A. was a holder of a fellowship

from CONICET. P.M.A.L. and A. K. are members of the Research Career of CONICET. N.L. P. is member of the Scientific Research Commission Prov. Buenos Aires (CIC).

References

1. Alonso MZ, Lucchesi PMA, Rodríguez EM, Parma AE, Padola NL. Enteropathogenic (EPEC) and Shigatoxigenic *Escherichia coli* (STEC) in broiler chickens and derived products at different retail stores. *Food Control*. 2012;23:351–5.
2. Chinen I, Epszteyn S, Melamed CL, Aguerre L, Martínez Espinosa E, Motter MM, Baschkier A, Manfredi E, Miliwebsky E, Rivas M. Shiga toxin-producing *Escherichia coli* O157 in beef and chicken burgers, and chicken carcasses in Buenos Aires, Argentina. *Int J Food Microbiol*. 2009;132:167–71.
3. EFSA Panel on Biological Hazards (BIOHAZ). Scientific opinion on VTEC-seropathotype and scientific criteria regarding pathogenicity assessment. *EFSA J*. 2013;11:3138.
4. Etoh Y, Murakami K, Ichihara S, Sera N, Hamasaki M, Takenaka S, Horikawa K, Kawano K, Takeishi T, Kuwana Y, Inoue A, Nagatsu Y, Hira Y, Takahashi M, Ito K. Isolation of Shiga toxin 2f-producing *Escherichia coli* (O115:HNM) from an adult symptomatic patient in Fukuoka Prefecture, Japan. *Jpn J Infect Dis*. 2009;62:315–7.
5. Fernández D, Irino K, Sanz ME, Padola NL, Parma AE. Characterization of Shiga toxin-producing *Escherichia coli* isolated from dairy cows in Argentina. *Lett Appl Microbiol*. 2010;51:377–82.
6. Friedrich AW, Bielaszewska M, Lan Zhang W, Pulz M, Kuczius T, Ammon A, Karch H. *Escherichia coli* harboring Shiga toxin 2 gene variants: frequency and association with clinical symptoms. *J Infect Dis*. 2002;185:74–84.
7. Galli L, Miliwebsky E, Irino K, Leotta G, Rivas M. Virulence profile comparison between LEE-negative Shiga toxin-producing *Escherichia coli* (STEC) strains isolated from cattle and humans. *Vet Microbiol*. 2010;143:307–13.
8. Gyles CL. Shiga toxin-producing *Escherichia coli*: an overview. *J Anim Sci*. 2007;85:E45–62.
9. Krüger A, Lucchesi PMA, Parma AE. Verotoxins in bovine and meat verotoxin-producing *Escherichia coli* isolates: type, number of variants, and relationship to cytotoxicity. *Appl Environ Microbiol*. 2011;77:73–9.
10. Llorente P, Barnech L, Irino K, Rumi MV, Bentancor A. Characterization of Shiga toxin-producing *Escherichia coli* isolated from ground beef collected in different socioeconomic strata markets in Buenos Aires, Argentina. *BioMed Res Int*. 2014:795104.
11. Padola NL, Sanz ME, Blanco JE, Blanco M, Blanco J, Etcheverria AI, Arroyo GH, Usera MA, Parma AE. Serotypes and virulence genes of Shigatoxigenic *Escherichia coli* (STEC) isolates from a feedlot in Argentina. *Vet Microbiol*. 2004;100:3–9.
12. Paton AW, Paton JC. Direct detection and characterization of Shiga toxigenic *Escherichia coli* by multiplex PCR for *stx1*, *stx2*, *eae*, *ehxA* and *saa*. *J Clin Microbiol*. 2002;40:271–4.
13. Reuben A, Treminio H, Arias ML, Chaves C. Presencia de *Escherichia coli* O157:H7, *Listeria monocytogenes* y *Salmonella* spp. en alimentos de origen animal en Costa Rica. *ALAN*. 2003;4:389–93.
14. Schmidt H, Scheef J, Morabito S, Caprioli A, Wieler LH, Karch H. A new Shiga-toxin variant (Stx2f) from *Escherichia coli* isolated from pigeons. *Appl Environ Microbiol*. 2000;66:1205–8.
15. Tutenel AV, Pierard D, Van Hoof J, Cornelis M, De Zutter L. Isolation and molecular characterization of *Escherichia coli* O157 isolated from cattle, pigs and chickens at slaughter. *Int J Food Microbiol*. 2003;84:63–9.