

Role of lactic acid bacteria in fermented vegetables

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SUMMARY: The consumption of fermented vegetables is widespread throughout the world and represents an important component of the human diet with considerable contribution to the food supply for a world population in continuous growth. Many of the fermented vegetables share a general process which requires salting and acidification steps. Among the microorganisms responsible for fermentation, lactic acid bacteria are the most relevant with important organoleptic, quality and safety benefits. This review deals with the microbial ecology of fermented vegetables focusing on the biodiversity of lactic acid bacteria, the most important molecular techniques used for their identification and genotyping, their importance for the formation of biofilms as well as their use as starter cultures for obtaining high-quality and safe vegetable products.

KEYWORDS: *Biofilms; LAB biodiversity; Multifunctional starters; Vegetables*

RESUMEN: *Papel de las bacterias del ácido láctico en verduras fermentadas.* El consumo de vegetales fermentados está muy extendido en el mundo y representa un componente importante de la dieta humana con un apoyo considerable a la cadena alimentaria para una población mundial en continuo crecimiento. Muchos de los vegetales fermentados comparten un proceso general, que requiere una puesta en salmuera y acidificación. Entre los microorganismos responsables de la fermentación, las bacterias del ácido láctico son las más relevantes con una importante influencia sobre aspectos organolépticos, de calidad y seguridad del producto final. Esta revisión trata sobre la ecología microbiana de los vegetales fermentados, prestando especial atención a la biodiversidad de las bacterias del ácido láctico, las técnicas moleculares más importantes utilizadas para su identificación y genotipado, su importancia para la formación de biofilms y su uso como cultivos iniciadores multifuncionales para la obtención de productos vegetales de alta calidad y seguridad.

PALABRAS CLAVE: *Biodiversidad de BAL; Biofilms; Cultivos iniciadores multifuncionales; Vegetales*

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1. INTRODUCTION

Fermented vegetables (cucumber, kimchi, sauerkraut, capers, carrots, table olives, etc.) play an important role in the human diet as a source of water-soluble vitamins, dietary fiber, phytosterols, phytochemicals and minerals (Gebbers, 2007). They also represent fundamental support for the feeding of a growing population. Fermentation is considered one of the oldest and cheapest methods used in food technology for food preservation. In addition, fermentation confers favorable effects to vegetables by improving the organoleptic characteristics of the final product (taste, color, texture, etc.), eliminates anti-nutritional components, prolongs shelf-life and increases the safety of the final products. The first reference of fermented vegetables was found in China during the construction of the Great Wall in III B.C. and constituted the basis of the workers' diet (Andersson, Daeschel and Eriksson, 1988).

Fermented vegetables can be considered both the edible part of the fruits (seeds and pulp) and other parts of the plant (leaves and roots). Many fermented vegetables share a common elaboration process which requires the use of salt and acidification by microorganisms (Figure 1). Sometimes a pre-step such as NaOH treatment, water washing, scalding, etc. is required, after harvesting the fruit. Pickles are prepared under three basic types of conditions: dry-salted, immersed in brine or non-salted cover solutions, where the fermentation takes place

during storage (Montet *et al.*, 2014). The development of certain autochthonous microorganisms present in the raw material, mainly lactic acid bacteria (LAB) and yeasts, are favored by the manufacturing of fermented vegetables (Di Cagno *et al.*, 2013; Arroyo-López, *et al.*, 2012b). The process of lactic fermentation consists of the conversion of sugars, mainly glucose, into lactic acid, which produces acidification at pH values below 4.6 units. This acidification inhibits the growth of undesirable spoilage and pathogenic microorganisms that can generate risks for public health.

2. THE MICROBIAL ECOLOGY OF FERMENTED VEGETABLES

The microorganisms present during vegetable fermentation are very diverse and may considerably affect the quality and safety of the final product. The microbiota initially present in the processes of lactic fermentation comes mainly from the fruit, although other elements such as brines, ingredients used and the industry's own environment influence its composition. The microbiota that colonizes the surface, and even the interior of the fresh fruit, is varied and depends on factors such as fruit maturity, climate and agricultural practices (Samish *et al.*, 1963; Mattos *et al.*, 2005). Mesophilic aerobic microorganisms on the surface of fresh pickles and cabbage can reach population levels of up to $5 \log_{10}$ CFU/mL (Pérez-Díaz *et al.*, 2015). This microbial

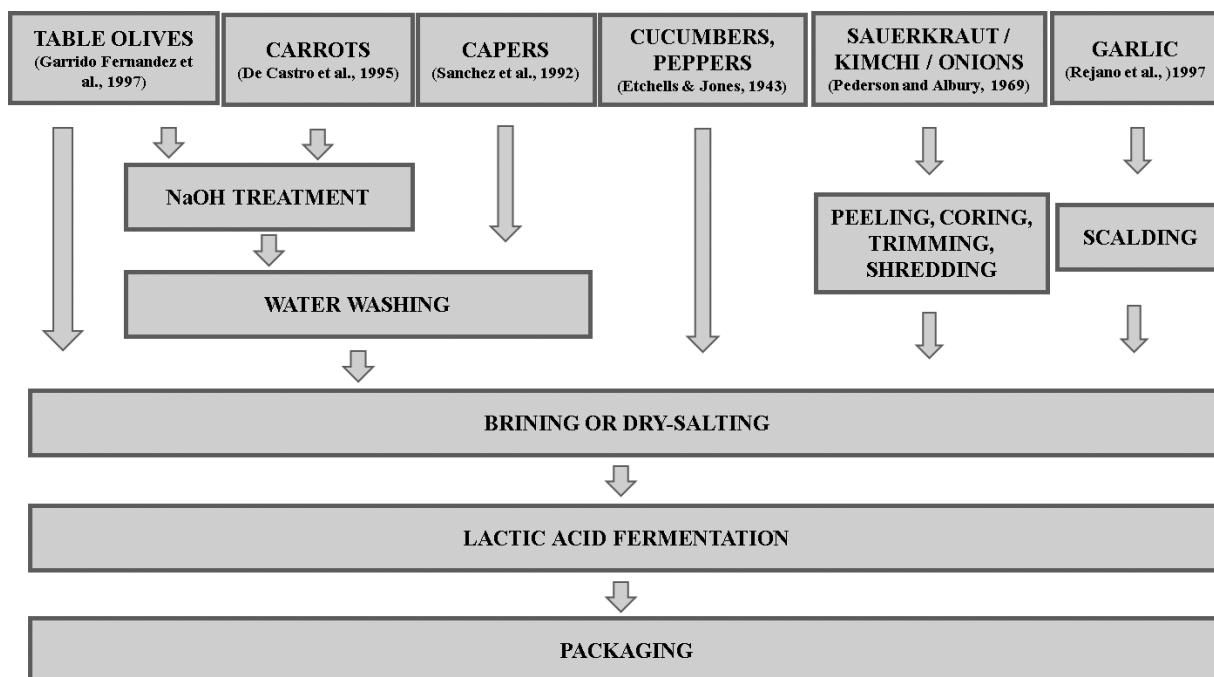


FIGURE 1. Flowchart of diverse fermented vegetable elaboration processes.

diversity is drastically reduced as the fermentation process evolves due to changes in the chemical conditions, mainly pH, acidity and salt concentration. A selection is made between the best adapted microbial groups and among them they compete for the nutrients and the dominance of the process. Enterobacteria, LAB and other groups of bacteria and yeasts can be active during the early stages of fermentation, depending on factors such as temperature, dissolved oxygen and the concentration of salts used in the brines. Subsequently, there is dominance by LAB (mainly *Lactobacillus*, *Pediococcus*, *Enterococcus* and *Leuconostoc* genera) throughout fermentation.

LAB are characterized by transforming fermentable material, mainly glucose and fructose, into organic acids through fermentation. The initial steps of vegetable fermentations are usually carried out by heterofermentative microorganisms with the production of lactic and acetic acids which make an important contribution to the flavor and aroma of the final product (Breidt *et al.*, 2013b). Then, they are replaced by the more acid-tolerant homofermentative microorganisms due to their capacity to produce lactic acid which induces a greater decrease in the pH but that inhibits the development of other microbial groups (Montet *et al.*, 2014). These pH changes, together with the concentration of salt, are responsible for obtaining stable and safe fermented vegetables. Lactic acid has advantageous preservative properties, which are responsible for fermented products having a longer shelf-life than their non-fermented counterparts. Also, other metabolites generated by LAB such as esters or bacteriocins contribute to the improvement of the organoleptic properties (acid and sour flavor) of the vegetable products produced by these processes. Fermentations carried out by LAB have also permitted the production of a large variety of foods with different aromas, flavors, and consistencies (Lücke, 1996).

In addition to the presence of LAB, yeast populations can also coexist during vegetable fermentation. In fact, they predominate in certain types of elaborations, such as directly brined olives (natural green and black) where the *Candida*, *Pichia*, *Debaryomyces*, and *Saccharomyces* genera are the most representative (Arroyo-López *et al.*, 2008; Botta and Coccolin, 2012). In the fermentation of gherkins, the genera *Torulopsis*, *Brettanomyces*, *Zygosaccharomyces*, *Hansenula*, *Torulasporea*, and *Kloeckera* also stand out (Etchells and Bell, 1950). Yeasts modulate the final organoleptic profile of the fermented product due to their capacity to produce volatile compounds associated with the development of flavor and aroma (Arroyo-López, *et al.*, 2012b; Hernández *et al.*, 2007; Rodríguez-Gómez *et al.*, 2010). However, yeasts can also be responsible for certain types of alterations. Excessive growth of

fermentative yeasts (*Saccharomyces cerevisiae* and *Wickerhamomyces anomalus*) could trigger an active production of CO₂ which damage the fruits with the formation of gas pockets or produce wiring in table olives (Vaughn *et al.*, 1972; Garrido-Fernández *et al.*, 1997). Certain yeasts, such as *Pichia manshurica* and *Issatchenkia occidentalis*, are associated with alterations in fermented vegetables due to their capacity to consume lactic acid and increase the pH of the product (Franco *et al.*, 2012). *Pichia kudriavzevii* is considered the main yeast responsible for the production of bad odor and flavor in kimchi, as well as softening (Moon *et al.*, 2014). Other fungi with pectinolytic capacity such as the genera *Alternaria*, *Fusarium* and *Mucor*, are responsible for the softening of pickles in brine (Costilow *et al.*, 1980).

In addition to LAB and yeasts, which are typically found in vegetable fermentations, gram-negative bacteria of the genera *Enterobacter*, *Citrobacter* and *Escherichia* are generally present at the beginning of fermentation. These microbial communities are inhibited because of lactic acid production by LAB, with a concomitant decrease in pH (Garrido-Fernández *et al.*, 1997). If the decrease in pH is not fast, Gram-negative bacteria can grow and produce CO₂ and compromise the quality and safety of the product leading to defects associated with the fruit, forming gas-pockets on the surface or inside the fruit, (“alambrado”) in gherkins and table olives (Fleming *et al.*, 1975; Garrido-Fernández *et al.*, 1997).

The presence of nutrients and a neutral or alkaline pH during the first stage of vegetable fermentation can contribute to the growth of *Clostridium* spp. These spoilage microorganisms can generate odors of decomposing organic matter or rancid butter, symptoms of putrid and butyric fermentations (Gililand and Vaughn, 1943). They are anaerobic spore-forming bacteria capable of colonizing the bottom of the fermenters where the concentration of oxygen is null. Sometimes, undesired secondary fermentation can be initiated by propionic acid bacteria such as *Propionibacterium* spp., converting sugars or lactic acid into propionic acid and CO₂, thus increasing the pH values (González-Cancho *et al.*, 1980). These conditions also enhance the growth of *Clostridium* species which, together with *Propionibacterium*, can promote the so-called “zapatería” alteration, giving off abnormal odors in table olives (Kawatomari and Vaughn, 1956; Plastourgos and Vaughn, 1957) or desirable aromas in sauerkraut (Babuchowski *et al.*, 1999). The resulting pH rise allows the growth of other microbes which are spoilage or pathogenic, compromising the safety of the product (Medina-Pradas and Arroyo-López, 2015). The presence of acetic acid bacteria and some strains of *Lactobacillus* spp. plays an important role in the onset of spoilage in fermented cucumbers by converting lactic acid into acetic acid (Johanningsmeier

and McFeeters, 2013; Medina *et al.*, 2016a). The increase in pH favors the growth of other species of *Propionibacterium* and *Pectinatus*. *Clostridium bifermentans* and *Enterobacter cloacae* which can metabolize the lactic acid and generate butyric and propionic acids, respectively (Breidt *et al.*, 2013a; Franco and Pérez-Díaz, 2013), and are responsible for the aroma of cheese and manure characteristic in spoiled fermented pickles.

3. BIODIVERSITY OF LAB IN FERMENTED VEGETABLES

The diversity of LAB species present in the different fermented vegetables that can be found on the market is wide, and responds to the different compositions presented by raw material, the environment and the physicochemical conditions prevailing during the fermentation process (Hurtado *et al.*, 2012).

In the specific case of table olives, the most representative genus, as mentioned above, is *Lactobacillus*, with *L. pentosus* and *L. plantarum* as the predominant species (Benítez-Cabello *et al.*, 2019; Botta and Cocolin, 2012), which show a great intra-specific diversity with the presence of different biotypes, depending on variety, type of processing, and geographical area. Other microorganisms identified during the fermentation process belong to the genera *Enterococcus*, *Pediococcus*, *Leuconostoc*, and *Lactococcus*, but always in a smaller proportion. Kimchi is also a fermented vegetable made with cabbages, radish, and various vegetables, and has great production and tradition in South Korea. As in the case of olives, there is also a great diversity of lactic bacteria, especially the genera *Weissella*, *Leuconostoc*, and *Pediococcus*. Furthermore, several

species of *Lactobacillus* and *Leuconostoc* have been identified in the fermentation of sauerkraut (cabbage). Finally, *P. ethanolidurans*, *E. thailandicus*, and different species of the genus *Lactobacillus* and *Leuconostoc* have been described as the most abundant species during the production of pickled cucumbers.

As a summary, Table 1 shows the main LAB species identified in diverse types of fermented vegetables.

4. IDENTIFICATION AND GENOTYPING OF LAB IN FERMENTED VEGETABLES: FROM MORPHOLOGICAL TECHNIQUES TO OMICS APPROACH

The conventional methods for the identification of LAB are based on morphological and physiological characteristics such as Gram stain, spore formation, enzyme production, and the determination of diverse biochemical reactions. Taking into account these techniques, the API system (BioMérieux, France) has been widely used. This biochemical methodology is based on the fermentative profile presented by microorganisms for a battery of carbohydrates. However, LAB present similar nutritional and growth needs, so these tests are not conclusive in most cases (Randazzo *et al.*, 2004).

At the end of the 90s, different molecular techniques were established, allowing for a more in-depth study of the bacterial ecology of food. The partial amplification by PCR (Polymerase Chain Reaction) of the DNA or RNA extracted directly from the microorganisms of interest, and its subsequent sequencing allows for microbial identification with much better precision thanks to the databases,

TABLE 1. Summary of LAB species identified in the main types of fermented vegetables

Vegetable Matrix	Specie	Reference
Table olives	<i>L. pentosus</i> , <i>L. plantarum</i> , <i>L. paraplantarum</i> , <i>L. parafarraginis</i> , <i>L. sanfranciscensis</i> , <i>Pediococcus</i> sp., <i>Lc. mesenteroides</i>	(Abriouel <i>et al.</i> , 2011; Hurtado <i>et al.</i> , 2012; Bautista-Gallego <i>et al.</i> , 2013; Benítez-Cabello <i>et al.</i> , 2016, 2019)
Kimchi	<i>L. curvatus</i> , <i>L. sakei</i> , <i>Lc. mesenteroides</i> , <i>Lc. gelidum</i> , <i>Lc. carnosum</i> , <i>Lc. gasicomitatum</i> , <i>P. pentosaceus</i> , <i>W. soli</i> , <i>W. cibaria</i> , <i>W. koreensis</i> , <i>W. cibaria</i>	(Jung <i>et al.</i> , 2013; Jang <i>et al.</i> , 2014; Hong <i>et al.</i> , 2015; Ji, Jang and Kim, 2015; Kyung <i>et al.</i> , 2015; Kim <i>et al.</i> , 2017)
Sauerkraut	<i>L. plantarum</i> , <i>L. pentosus</i> , <i>Lc. mesenteroides</i> , <i>L. brevis</i> , <i>L. sakei</i> , <i>L. curvatus</i> , <i>L. paraplantarum</i> , <i>L. coryniformis</i> , <i>P. pentosaceus</i> , <i>Lc. citreum</i> , <i>Lc. argentinum</i> , <i>Weissella</i> sp.	(Johanningsmeier <i>et al.</i> , 2007; Plengvidhya <i>et al.</i> , 2007; Qing Yue <i>et al.</i> , 2013; Yan <i>et al.</i> , 2015)
Cucumbers	<i>L. pentosus</i> , <i>L. plantarum</i> , <i>L. brevis</i> , <i>L. paracasei</i> ; <i>Weissella</i> spp., <i>P. ethanolidurans</i> , <i>Leuconostoc</i> spp., <i>Lactococcus</i> spp	(Breidt <i>et al.</i> , 2013a; Medina <i>et al.</i> , 2016a; Pérez-Díaz <i>et al.</i> , 2016)
Other fermented vegetables	<i>Enterococcus thailandicus</i> , <i>E. casseliflavus</i> , <i>Lc. lactis</i> , <i>Lc. mesenteroides</i> , <i>W. hellenica</i> , <i>L. pentosus</i> , <i>L. plantarum</i> , <i>L. paraplantarum</i> , <i>L. brevis</i> , <i>L. citreum</i> , <i>L. alimentarius</i> , <i>L. paracasei</i> , <i>L. buchneri</i> , <i>P. ethanolidurans</i>	(Breidt <i>et al.</i> , 2013b; Tamminen <i>et al.</i> , 2004; Chen <i>et al.</i> , 2012; Yu <i>et al.</i> , 2012; Wouters <i>et al.</i> , 2013; Elmaci <i>et al.</i> , 2015; Reina <i>et al.</i> , 2015)

such as NCBI GenBank (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>). Other molecular techniques use random amplified polymorphic DNA (RAPD), repetitive elements of bacterial DNA (rep-PCR) or combined PCR followed by enzymatic digestion (RFLP). These techniques create fragments with different lengths, giving rise to a specific band profile for each strain (finger printing).

In recent years, many studies have used this molecular approach to distinguish different biotypes within the same species in the field of fermented vegetables (De Bellis *et al.*, 2010; Franzetti *et al.*, 2011; Aponte *et al.*, 2012; Breidt *et al.*, 2013a; Benítez-Cabello *et al.*, 2019). These molecular tools are very popular to characterize and carry out phylogenetic studies on microbial communities (Abriouel *et al.*, 2012; Lucena-Padrós *et al.*, 2014; Romero-Gil *et al.*, 2016).

Due to the disadvantages of these culture-dependent techniques, the term “independent culture techniques” was defined, without the need for the culture of microorganisms in which DNA or RNA are extracted directly and analyzed from the food matrix. Undoubtedly, the independent culture methods offer several advantages over the dependent culture methods, such as: i) to avoid the use of specific culture media; ii) they are based on the presence of DNA, RNA, or proteins; iii) the physiological state of the cell has no affect; and iv) these methods are able to detect populations whose concentration is lower than the detection limits of traditional methods (Cocolin and Ercolini, 2007). However, the selection of the target for these techniques must satisfy two premises: i) to be common for all members of the microbial group to consider; and ii) the presence of conserved regions for the universal primer design, and variable regions for a possible differentiation. Clear examples are the genes encoding ribosomal RNA (rRNA), such as various regions of the 16S genes for bacteria, or 26S and ITS genes for yeasts.

At the end of the 90s, the DGGE technique (gel electrophoresis with denaturation gradient) was introduced in food microbiology (Ampe *et al.*, 1999). Amplification of the variable 16S region combined with DGGE allowed for the discrimination of the PCR products based on their mobility as determined by the specific DNA sequence of the amplicon. DGGE has made it possible to better understand the microbial diversity of a wide variety of foods, including kimchi (Lee *et al.*, 2005) and table olives (Abriouel *et al.*, 2011; Benítez-Cabello *et al.*, 2016; Lucena-Padrós *et al.*, 2015).

Finally, from the 2010s, the use of next-generation sequencing (NGS) and metagenomics in foods, including fermented vegetables, has become widespread. These novel techniques have revolutionized the field of microbial ecology in food through more precise identification of microbial taxa without the

need for culture-dependent methods. In the particular case of fermented vegetables, metagenomics has become an ideal tool for the study of the bacterial biodiversity of table olives (Cocolin *et al.*, 2013; De Angelis *et al.*, 2015; Medina *et al.*, 2016b; 2018; De Castro *et al.*, 2018), cucumbers (Medina *et al.*, 2016a), kimchi (Hong *et al.*, 2015, Kyung *et al.*, 2015) and other fermented vegetables (Reina *et al.*, 2015). However, these studies based on massive sequencing can generate partial representations of microbial diversity, so Ferrocino and Cocoli (2017) proposed using a multi-omic approach in the future, for example, combining it with metatranscriptomic or metaproteomics data.

5. APPLICATION OF LAB AS STARTER CULTURES IN FERMENTED VEGETABLES

The fermentation processes will be defined by different physicochemical parameters (salinity, acidity, temperature, presence of antimicrobial compounds, etc.) which will be decisive for obtaining a fermented product of high quality, which is safe and microbiologically stable. Fermentation can occur spontaneously; however, several authors recommend the use of starter cultures in order to manage the process (Corsetti *et al.*, 2012; Lee *et al.*, 2015). For many years, the search for starters with application in vegetables has been practically strictly focused on the activity of LAB and their technological applications. Recently, several authors have emphasized the role of selected yeasts in combination with LAB during processing due to their multifunctional features (Arroyo-López *et al.*, 2012a). Yeasts could be especially effective in diverse fermented vegetables where LAB are partially inhibited by the presence of high concentrations of antimicrobial compounds, such as directly brined table olives (Arroyo-López *et al.*, 2012a; Ruiz-Barba *et al.*, 1993).

The selection criteria for LAB for their use as starter cultures has traditionally been related to the homo-fermentative metabolism that implies a rapid rate of sugar consumption and lactic acid production, good adaptation to intrinsic conditions of temperature, pH, salt, and inhibitory compounds throughout the process, and resistance to bacteriophages, bacteriocin production as an important factor in establishing strains which help to increase the quality and safety, improvement in organoleptic characteristics, minimum nutritional requirement, enzymatic activities (esterase and β -glucosidase) related to the biological de-bittering of fruits, and good imposition rates, among many other characteristics (Benítez-Cabello *et al.*, 2019; Hurtado *et al.*, 2012). Technologically, a starter culture must have the ability to prevail against the autochthonous microbiota and resist freezing or freeze-drying processes for producing a commercial starter. Many of these starter cultures currently used in fermented

vegetables belong to the *Lactobacillus*, *Leuconostoc*, and *Pediococcus* genera. Furthermore, these starter cultures should be also monitored to avoid possible negative characteristics, such as production of biogenic amines, off flavor and odor, pectinolytic activity, etc. The presence of biogenic amines has been reported in sauerkraut and table olives, where the presence of amino-biogenic spoilage microorganisms can result in high putrescine, cadaverine and tyramine contents (Medina-Pradas and Arroyo-López, 2015; Rabie *et al.*, 2011).

Probiotic foods have been associated for a long time with dairy products with the disadvantage of not being able to be consumed by people intolerant to lactose, in addition to their high cholesterol content (Granato *et al.*, 2010). This scene opens a new possibility for the development of starter cultures in fermented vegetables with probiotic potential. Many researchers have studied the probiotic characteristics of several LAB species from the fermentation of sauerkraut, kimchi, cabbage, carrot or fresh beans (Argyri *et al.*, 2013; Botta *et al.*, 2014; Benítez-Cabello *et al.*, 2019; Peres *et al.*, 2012; Di Cagno *et al.*, 2013;). Studies with LAB probiotic strains isolated from artichokes and table olives showed similar, even higher survival rates than those from milk probiotics during simulated human digestion (Bautista-Gallego *et al.*, 2013; Lavermicocca *et al.*, 2005; Arroyo-López *et al.*, 2014).

Therefore, new challenges for the development of starter cultures should be focused on the study of the probiotic characteristics of autochthonous microorganisms present in vegetables, highlighting LAB and yeast, with the aim of establishing multifunctional mixed starter cultures that are complementary both in their properties and modes of action. Besides an appropriate technological behavior, a multifunctional starter culture must be able to exert other biological activities of interest such as cholesterol removal, inhibitory or exclusion activity against pathogenic microorganisms, resistance to acidic conditions of gastric and pancreatic digestion, phytase, antioxidant, and lactase activities, no antibiotic resistance, production of functional exopolysaccharides, adhesion to human cellular lines, and immunomodulatory activity, among others.

6. LAB-FORMING BIOFILMS IN FERMENTED VEGETABLES

In recent years, diverse fermented vegetables have been proven as carriers of potential probiotic microorganisms to the human body. Several studies with scanning electron microscopy have observed that some microorganisms present in the fermentation of vegetables have the ability to form biofilms on the surface of the food, as occurs with table olives (Arroyo-López *et al.*, 2012a; Domínguez-Manzano *et al.*, 2012). This biofilm is composed

of a poly-microbial community, mostly of lactobacilli and yeasts, which has been embedded by an extracellular exopolysaccharide matrix whose function is to protect them from the environment and bind them to the epidermis of the fruit thanks to its adhesive properties. The EPS produced by these biofilm-forming microorganisms also has functional properties, such as to avoid adhesion of other pathogen microorganisms to animal cellular lines (González Ortiz *et al.*, 2013). Biofilms can be formed on biotic (vegetables) or abiotic (wall fermentation vessels, machinery, etc.) surfaces.

The study of the interactions between yeasts and LAB to form biofilms, the development of mixed starter cultures and the formation of biofilms is quite recent in the field of fermented vegetables. It opens the possibility of converting these foods into an excellent vector of beneficial microorganisms to the final consumer. In several studies conducted on table olive biofilms, mixed microbial populations have been found of up to $8 \log_{10}$ CFU g⁻¹ of *L. pentosus* among LAB, and *Pichia galeiformis*, *Candida sorbosa*, *Geotrichum candidum*, *S. cerevisiae* and *W. anomalus* among yeasts (Arroyo-López *et al.*, 2012a; Domínguez-Manzano *et al.*, 2012). Moreover, these microorganisms did not lose viability during long-term storage in olive packing at room temperature without a cold chain (Rodríguez-Gómez *et al.*, 2014). Certain combinations of yeasts and lactic acid bacteria favor the formation of biofilms. León-Romero *et al.*, (2016) reported that the interaction between diverse genotypes of *C. boidinii* and *W. anomalus* in a mixed culture with *L. pentosus* formed the best biofilm, which was not found for other strains. Biofilm formation can be inhibited by the presence of D-(+) mannose and stimulated even in the absence of cell-cell contact between yeast and LAB species.

However, the research on the genes involved in a biofilm formation process, the modulation of their expression in a mixed culture, the interaction between yeast-LAB and with the surrounding matrix is still scarce, despite the importance of yeast-LAB cultures and the presence of biofilms in the elaboration of fermented vegetables.

7. CONCLUSIONS

In the last decade, research on fermented vegetables has been building on new and advanced techniques that have allowed for establishing a more solid base of knowledge about the microbial ecology implied in elaboration processes. In particular, the molecular and metagenomic techniques and the further bio-informatic analysis will allow for an in-depth study of the role of LAB throughout fermentation, the microbial changes during processing conditions, geographical and varietal influences, and application of cleaning procedures, etc.

Moreover, the use of a new generation of multifunctional starter cultures (technological + probiotic) will result in better process control, reducing economic losses and spoilage, improving the quality and safety aspects, but at the same time, producing a product with higher functional value open new market niches.

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